

# Nanotechnology Applications for Glioblastoma

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## KEYWORDS

- Malignant brain tumors • Glioblastoma • Magnetic nanoparticles • Nanoparticles
- Convection-enhanced delivery • MRI • EGFR • Thermotherapy

## KEY POINTS

- GBM remains a difficult tumor to treat due to its infiltrative nature.
- Nanoparticles present a new way to target infiltrating cells.
- Magnetic nanoparticles (MNPs) can be used as MRI contrast agents as well as therapeutic agents by the use of thermotherapy.
- In a nanoparticle formulation chemotherapeutics can be more efficacious than conventional chemotherapeutic agents due to their ability to target GBM cells and release drug.
- Gene delivery through the use of nanoparticles may be a safe option to deliver therapeutic genes to tumor cells.
- Brachytherapy delivered by radioactive nanoparticles can provide long-term focused radiation therapy to these lesions.
- Gold nanoparticles can be used to treat tumors through phototherapy, where deep penetrating near-infrared light can be used to inhibit tumor growth.
- Nanoparticles can be delivered safely systemically or by bulk flow using convection-enhanced delivery (CED) directly to the tumor.
- Magnetic targeting can be used to enhance the delivery of MNPs by directing the delivered particles to the area of interest.

## INTRODUCTION

Glioblastoma (GBM) is the most common primary malignancy of the brain as well as its most malignant.<sup>1</sup> The median survival after radiation and chemotherapy ranges from 12 to 15 months, despite advances in surgery, radiation, and chemotherapy.<sup>2</sup> GBM tumors are nearly uniformly fatal due to local recurrence.<sup>3–5</sup> Even for lesions amenable to gross

surgical resection, infiltrating cancer cells beyond the boundaries of the enhancing lesion are responsible for tumor recurrence as well as radiation and chemotherapy resistance.<sup>6,7</sup>

Cancer nanotechnology has recently emerged as a field that may provide answers to some of the difficulties encountered in treating GBM. Nanoparticles, defined as particles less than 100 nm in hydrodynamic size, have been used in the

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treatment of various cancers.<sup>8</sup> The use of biocompatible nanomaterials has permitted the fabrication of nanoparticles with capabilities that surpass those of conventional agents. Chemotherapy-loaded nanoparticles have resulted in sustained-release formulations that can lower systemic toxicity and produce greater antitumor effects. Recently developed nanoparticles can cross the blood-brain barrier (BBB) after systemic administration or be distributed in the brain by CED to target GBM cells therapeutically while harboring elements that may enable imaging of the particle and the target. The field has been moving at a rapid pace, enabling nanoparticles to be used in recent clinical trials.<sup>9</sup> Although not exhaustive, the list of nanoparticles used in the treatment of experimental GBM includes polymeric particles, micelles,<sup>10</sup> nanoshells,<sup>11</sup> quantum dots,<sup>12</sup> and magnetic iron oxide nanoparticles (IONPs).<sup>13</sup> Nanotubes are another formulation of nanoparticle, used to create structures that can trap diagnostic or therapeutic modalities within a cage. This article discusses the use of different nanoparticle formulations in strategies to image and treat GBM, including delivery schemes.

## MAGNETIC NANOPARTICLES

### *MRI Contrast Properties of MNPs*

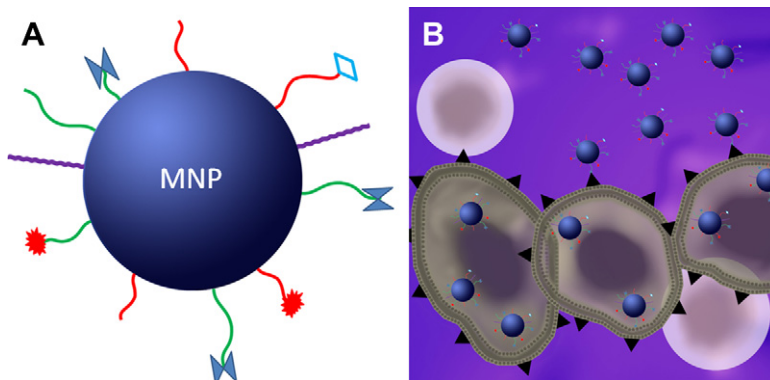
The base of the promise for theranostic nanoparticles with both therapeutic and diagnostic ability hinges on the idea that such nanoparticles will be able to image where the lesion is and treat it. MNPs have attracted particular interest in this respect due to their unique paramagnetic properties that enable their detection by MRI.<sup>14,15</sup> These MNPs have shown great potential as T<sub>1</sub> or T<sub>2</sub> contrast agents in MRI,<sup>16,17</sup> with superparamagnetic iron oxide-based nanoparticles (SPIOs) as

the most commonly investigated type of MRI contrast agents.<sup>18</sup> Since 1990, ultrasmall SPIOs (USPIOs), smaller than 50 nm, have been considered an MRI contrast agent,<sup>19</sup> and most of the MRI data regarding nanoparticles references these particles. USPIOs can be visualized in T<sub>2</sub>-weighted MRI sequences (T<sub>2</sub> contrast agents) as a hypointense (dark) signal (negative contrast enhancement) or with T<sub>1</sub>-weighted MRI sequences (T<sub>1</sub> contrast agents) as a hyperintense (bright) signal (positive contrast enhancement).<sup>20–22</sup>

USPIOs can provide contrast for a longer period of time<sup>23</sup> compared with gadolinium (Gd)-based contrast agents that are rapidly eliminated by the kidney.<sup>24,25</sup> USPIOs are also taken up by tumor cells as well as by reactive phagocytic cells (eg, microglia) found in brain tumors. The USPIOs can reside within brain tumors much longer than Gd-based agents, with a peak enhancement noted at 24 to 28 hours and persisting up to 72 hours after administration.<sup>26,27</sup> These agents may provide a safe alternative for patients at risk for nephrogenic systemic fibrosis, because preliminary studies have shown no adverse renal effects.<sup>27,28</sup>

### *MNPs for Targeted Brain Tumor Imaging*

Targeting of tumor cells can increase the benefits provided by nanoparticles as contrast agents. IONPs are taken up by GBM cells both in vivo and in vitro.<sup>29,30</sup> Surface functionalization further enhances tumor uptake of these particles.<sup>31</sup> Tumor-specific ligands conjugated to MNPs can further enhance the uptake within targeted tumor tissue (**Fig. 1**).<sup>32,33</sup> Antibodies, peptides (including toxins), cytokines, and chemotherapeutic agents have been reported as possible MNP ligands.<sup>34</sup> Amphiphilic triblock copolymer IONPs can be conjugated with a purified antibody that selectively



**Fig. 1.** Theranostic MNPs and tumor targeting. (A) Illustration of an MNP with different functional groups on the surface, which permit molecular targeting, imaging, enhanced plasma circulation times, and/or therapy. (B) Illustration of MNPs functionalized with tumor cell-specific ligands binding cancer cells (*large irregular cells*) instead of normal cells (*pink*). Internalization of MNPs is shown in cancer cells as well.

binds to the epidermal growth factor receptor (EGFR) deletion mutant, EGFR vIII, which is solely expressed by a population of GBM tumors.<sup>35</sup> Such nanoparticles exhibit MR contrast enhancement of GBM cells and can target these therapy-resistant cancer cells in vitro and in vivo.

Chlorotoxin, derived from scorpion venom, specifically binds to matrix metalloproteinase 2 (MMP-2), which is overexpressed on the surface of GBM cells.<sup>36,37</sup> MMP-2 degrades the extracellular matrix during tumor invasion, and chlorotoxin can be used to bind the MMP-2 and inhibit infiltration.<sup>38,39</sup> Chlorotoxin conjugated to MNPs can act as MRI contrast agents and the addition of a Cy5.5 molecule makes these suitable for use as an intraoperative fluorescent dye as well.<sup>40–42</sup>

F3 is a small peptide that specifically binds to nucleolin overexpressed on proliferating endothelial cells of tumor cells and the associated vasculature.<sup>43</sup> F3-coated IONPs can provide significant MRI contrast enhancement of intracranial rat-implanted tumors, compared with noncoated F3 nanoparticles, when administered intravenously.<sup>44</sup>

A molecular MRI contrast agent, consisting of SPIO coated with dextran, was functionalized with an anti-insulinlike growth factor binding protein 7 (anti-IGFBP7) single-domain antibody and was found by both MRI and in vivo fluorescent imaging to target the vasculature of GBM cells.<sup>45</sup>

Gd has also been incorporated into some therapeutic nanoparticles to enable them to be tracked using MRI. One group has designed nanoparticles containing Gd, which are rapidly taken up by the GL261 tumor cell line and show MRI contrast when these cells are then cultured in a chick embryo host.<sup>46</sup> Gd nanoparticles functionalized with diethylenetriaminepentaacetic acid (DTPA) can also be used as a radiosensitizing agent.<sup>47</sup> Fullerene magnetic nanotubes have been made such that Gd can be trapped within these structures to make them an effective contrast agent, along with whatever therapeutic modality is also associated with the fullerene cage.<sup>48,49</sup> It is also possible to internalize IONPs in these larger nanotube structures so that the magnetic properties of iron oxide can be used, allowing clinicians to localize these particles to a particular area. This, together with surface targeting, can greatly increase the amount of intake and resultant therapeutic effect of these particles.<sup>50</sup>

### ***MNPs for Optical Delineation of Brain Tumors***

Although surgical intervention is not curative in GBM, obtaining a maximal resection is important for survival.<sup>51</sup> The use of intraoperative MRI and neuronavigation has increased extent of resection

and outcome.<sup>52–54</sup> Recently, fluorescence-guided surgery after oral administration of 5-aminolevulinic acid (5-ALA) has resulted in more complete resection of malignant gliomas.<sup>55,56</sup> Laboratory studies have attempted to find ways to use optical aides to increase the contrast between normal and tumor tissue,<sup>57–59</sup> and these methods have shown improvement in the extent of tumor resection in clinical use.<sup>60,61</sup>

Fluorescent molecules have already been successfully incorporated into several nanoparticles. An IONP-Cy5.5 molecule has been used in many preclinical studies,<sup>40,41,62</sup> giving it the dual benefits of MRI detection and possibly enhanced surgical contrast using the fluorescent properties of the particle. This also could lead to theranostic particles that could be injected preoperatively to outline malignant tissue that would need to be resected at surgery.

### ***MNPs for Stem Cell Tracking***

The ability of MNPs to act as MRI contrast agents can be used to track stem cell tropism to malignant brain tumors in vivo. Intracranially administered neural stem cells have tropism for GBM tumors, making them attractive for tumor-targeting gene therapy.<sup>63–65</sup> Mesenchymal stem cells have also been found to migrate to tumor cells.<sup>66</sup> By labeling these cells with IONPs, this migration can be visualized on MRI.<sup>67,68</sup> Magnetically labeled hematopoietic stem cells can also be tracked to gliomas in this fashion.<sup>69</sup>

### ***MNPs for Thermotherapy of GBM***

One of the more unique features of MNPs is the ability to induce hyperthermia when exposed to alternating magnetic fields. Temperature elevations in the range of 41°C to 46°C can cause cells to undergo heat stress, resulting in protein denaturation, protein folding, aggregation, and DNA cross-linking.<sup>70</sup> This process can induce apoptosis and heat shock protein expression. At the tissue level, moderate hyperthermia causes changes in pH, perfusion, and oxygenation of the tumor microenvironment.<sup>71–74</sup> These effects, combined with chemotherapy and radiation, can have a synergistic effect.<sup>74–78</sup>

Hyperthermia can be induced in MNPs through the use of an appropriate alternating magnetic field of the right amplitude and frequency to heat up the nanoparticles. A predictable and sufficient amount of heat known as the specific absorption rate is produced. The MNPs use several different mechanisms to convert the magnetic energy into heat energy. Néel relaxation is caused by rapidly occurring changes in the direction of magnetic

moments relative to crystal lattice. Brownian relaxation results from the physical rotation of MNPs within the medium in which they are placed. Both internal (Néel) and external (brownian) sources of friction lead to a phase lag between applied magnetic field and the direction of magnetic moment, producing thermal losses (Fig. 2).

MNPs can be specifically engineered to maximize their suitability for hyperthermia by producing greater saturation magnetization, optimal anisotropy, and larger size within the constraints of nanoparticle production.<sup>79–81</sup> MNPs suitable for thermotherapy can be made from a combination of various metals, including manganese (Mn), iron (Fe), cobalt (Co), nickel (Ni), zinc (Zn), and magnesium (Mg) and their oxides.<sup>82–89</sup> Ferrites of the various metals are frequently used in these settings, such as cobalt ferrites ( $\text{CoFe}_2\text{O}_4$ ), manganese ferrites ( $\text{MnFe}_2\text{O}_4$ ), nickel ferrites ( $\text{NiFe}_2\text{O}_4$ ), lithium ferrites ( $\text{Li}_{0.5}\text{Fe}_{2.5}\text{O}_4$ ), mixed ferrites of nickel–zinc–copper, and cobalt–nickel ferrites.<sup>85–91</sup> There are also ferromagnetic nanoparticles that are iron based and have greater magnetic properties than IONPs.<sup>79</sup> These cobalt ferrites–based nanoparticles produce greater hyperthermia effects at much lower concentrations than IONPs. FeNPs are comprised of an iron core surrounded by an iron oxide layer to permit stability. Nevertheless, owing to their lack of toxicity, excellent biocompatibility, and their capacity to be metabolized,<sup>92–94</sup> iron oxide–based MNPs are actively being studied for thermotherapy of brain tumors.

MNP-based hyperthermia has been evaluated for feasibility in animal models and in human patients with malignant brain tumors. Dextran-coated or aminosilane-coated IONPs have been used for thermotherapy in a rodent GBM model<sup>95</sup> and in a human clinical trial in patients with recurrent GBM.<sup>9,96</sup> Intratumoral injection of aminosilane-

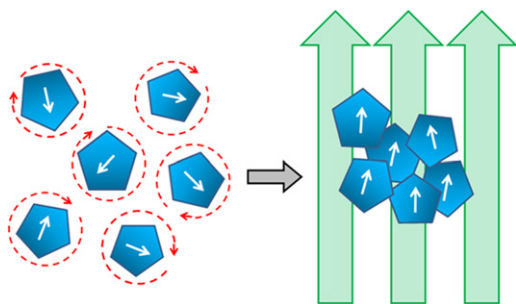
coated IONPs (core size 12 nm) and application of an alternating magnetic field (100 kHz) in several sessions before and after adjuvant fractionated radiation therapy was given. With a high concentration of IONPs (>100 mg/mL), this achieved effective thermotherapy with a median peak temperature within the tumor of 51.2°C. This phase II clinical trial successfully demonstrated safety and efficacy of thermotherapy of malignant brain tumors with MNPs in humans, with a significant increase in overall survival compared with a reference population. Further randomized studies will be required to validate the promise of this treatment modality.

## NANOPARTICULARIZED CHEMOTHERAPEUTIC AGENTS

Although few conventional chemotherapeutics have been proved effective in GBM, chemotherapeutics in a nanoparticle formulation offer possible advantages. These often can be targeted, evade the reticuloendothelial system for prolonged circulatory time, and potentially cross the BBB better than standard chemotherapy agents. Polyethylene glycol (PEG)-coated paclitaxel (taxol) nanoparticles have been shown to offer superior bioavailability compared with free paclitaxel with a survival advantage shown in a rodent glioma model.<sup>97</sup> Poly(D,L-lactide-co-glycolide) (PLGA) nanoparticles are another form of biocompatible nanoparticles. CED of these nanoparticles, loaded with camptothecin, has been shown efficacious in a rodent glioma model.<sup>98</sup> Although the controlled release offered by nanoparticles can reduce systemic toxicity and allow drug to be slowly released only when it has reached its target, there is also a need to ensure that an adequate dose is delivered to the lesion being treated. Nanoparticles have been developed that are thermosensitive, releasing their drug preferentially when the temperature has been increased.<sup>99</sup> When delivered with gold nanorods, concurrent photothermal hyperthermia can release the drug from the heat sensitive nanoparticle, thus increasing efficacy.

## GENE DELIVERY WITH NANOPARTICLES

The Cancer Genome Atlas has revealed the multiple genetic aberrations in GBM tumors that can serve as therapeutic targets provide targets.<sup>100</sup> Cationic solid lipid nanoparticles can be conjugated to PEGylated therapeutic *c-Met* small interfering RNA and reduce human GBM tumor growth in a rodent model without significant toxicity.<sup>101</sup> Another nanoparticle, containing the integrin-binding motif RGD, together with the PEG-polyethylenimine (PEI) nonviral gene carrying



**Fig. 2.** MNP response to alternating magnetic fields and thermotherapy. Application of applied magnetic fields (arrows) orients the MNPs on the right from their random orientation on the left in the absence of magnetic fields. Random orientation on the left produces thermal losses, allowing for hyperthermia generation by the MNPs.

nanoparticle, was able to deliver a plasmid expressing the tumor necrosis factor-related apoptosis-inducing ligand with increased efficiency and increase survival in a rodent glioma model.<sup>102</sup>

## NANOPARTICLES FOR BRACHYTHERAPY

Brachytherapy, where localized radiotherapy is delivered directly to a tumor, has been explored as a strategy with nanoparticles. In an orthotopic xenograft brain tumor model, a functionalized fullerene nanoparticle (<sup>177</sup>Lu-DOTA-f-Gd<sub>3</sub>N@C<sub>80</sub>), with radiolabeled lutetium 177 (<sup>177</sup>Lu) and tetraazacyclododecane tetraacetic acid (DOTA), provided an anchor to deliver effective brachytherapy and longitudinal imaging of the tumor.<sup>103</sup> Internal fractionated radiation has also been achieved using a lipid nanoparticle formulation of radionuclides, such as <sup>188</sup>Re-SSS in the 9L rat glioma cell line.<sup>75</sup>

## GOLD NANOPARTICLE PHOTOTHERAPY

Gold nanoparticles can be designed as nanoshells, consisting of a spherical dielectric core nanoparticle surrounded by thin sheet metal.<sup>76</sup> The size of each layer of the nanoshell can be tailored to enable it to have a peak light absorption at 800 nm, in the near infrared range. Light in this region of the electromagnetic spectrum has minimal absorption by water and biologic chromophores, allowing it to pass deep into tissues without losing much of its energy. This region of the electromagnetic spectrum is notable for minimal absorption by water and biologic chromophores. Thus, light of this wavelength may penetrate deep into tissues with minimal disruption. This has enabled researchers to produce these gold nanoparticles, which can be activated by light and kill GBM cells in vitro.<sup>77</sup> One group has used macrophages loaded with gold nanoshells to deliver these particles to glioma spheroids to then be activated by near infrared light, inhibiting growth.<sup>78</sup>

## MALIGNANT BRAIN TUMOR DELIVERY OF NANOPARTICLES

Delivery of therapeutic agents to GBM tumors remains a formidable challenge. Systemic delivery is limited by the BBB, nonspecific uptake, nontargeted distribution, and systemic toxicity. The benefits and drawbacks of the use of systemic delivery, systemic delivery augmented by magnetic targeting, and direct infusion in the brain known as CED are examined.

### Systemic Delivery

The reticuloendothelial system can significantly reduce the amount of nanoparticle available to treat

the lesion by nonspecific uptake in the liver, kidney, spleen, and circulating macrophages.<sup>104,105</sup> This can be addressed by biocompatible surface coating of nanoparticles, which can increase their circulation time.<sup>106</sup> The BBB further obstructs delivery by preventing the entry of most particles from the circulation into the interstitial space of the brain. It is well known that the vasculature in GBM, however, is not phenotypically normal, due to open endothelial gaps and atypical angiogenesis, allowing more efflux of intravascular material into the tumor mass.<sup>107–109</sup> The enhanced permeability and retention effect is used to describe the selective extravasation of macromolecules into the tumor interstitium through the hyperpermeable tumor vasculature.<sup>110</sup> By attaching tumor-specific targeting ligands, delivery has been shown to be increased in a rodent model, because the extravasated treatment is more likely to be taken up by the lesion.<sup>44,111</sup>

Integrins are overexpressed in GBM at the brain tumor border, and one of the integrin-binding motifs is RGD. Conjugating this peptide to PEG and PEI creates a nanoparticle, which was targeted to GBM and found to prolong survival in rodents implanted with human intracranial GBM xenografts.<sup>102</sup> Meng and colleagues used PEI conjugated to DNA and myristic acid, a hydrophobic molecule, can enhance the ability of the PEI/DNA complexed nanoparticles to cross the BBB, thus showing a treatment effect in GBM tumor models.<sup>112</sup>

PLGA nanoparticles have been shown to cross the BBB. The use of surfactants, such as poloxamer 188 (Pluronic F-68) or polysorbate 80 (Tween 80), can enhance the transport of the particles and increase the delivery of drugs conjugated to them and increase intracellular uptake.<sup>113–115</sup> A recent study demonstrated that conjugating transferrin, a protein known to be actively transported across the BBB, enhances the delivery of these particles to the brain, with an intact BBB as well as a disrupted BBB with an intracranial lesion.<sup>116</sup>

The  $\alpha$ -helical amphipathic peptide D[KLAKLAK]<sub>2</sub> was originally designed as a synthetic antibacterial peptide that disrupts the bacterial cell membrane but is less toxic to eukaryotic cells. When conjugated to a mitochondrial peptide, CGKRK, IONP-derived nanoworms (due to their elongated shape), these particles localize to the mitochondria of tumor cells and cure tumors in a rodent tumor model. The nanoparticles could be seen to localize to the tumor on MRI.<sup>111</sup>

### Magnetic Targeting

The concept of magnetic targeting of malignant brain tumors has also been demonstrated in



preclinical rodent models<sup>117,118</sup> as a method to enhance the systemic delivery of MNPs to malignant brain tumors. By using a magnetic field targeted to the region of interest, it has been shown that delivery of MNPs can be increased over the delivery to lesions when a magnetic field is not used.<sup>119</sup> There are concerns in how efficacious the translation of this technique will be to human studies, because the depth of the lesions in the human brain limit the ability to precisely target a lesion with a magnetic field.<sup>118</sup> Nevertheless, this remains an area for increased study.

In an effort to enhance the delivery and deposition of MNPs into malignant brain tumors, many studies have examined using strategies to open the BBB. Focal ultrasound represents a noninvasive technique, which can selectively disrupt the BBB and increase the enhanced permeability and retention effect in a targeted region of the brain.<sup>120–122</sup> Focal ultrasound and magnetic targeting have been used synergistically to enhance the delivery and the deposition of chemotherapy (epirubicin)-loaded MNPs into tumor-bearing animals. Epirubicin delivery and brain tumor accumulation was significantly enhanced by the combined focal ultrasound/magnetic targeting approach of epirubicin-MNPs.<sup>123</sup>

### **Convection-Enhanced Delivery**

CED, where bulk flow is used to distribute infusate throughout the brain with a pressure gradient, is a well-established technique for delivery of molecules to the brain.<sup>124</sup> CED bypasses the BBB, allowing targeted delivery of infusate to the parenchyma of a region of interest through a catheter. A pump is connected to each infusion catheter to ensure a positive pressure gradient during delivery for convection of molecules through the interstitium of the brain. The pressure gradient created by the pump greatly augments the delivery that would be achieved by the use of simple diffusion alone.<sup>125</sup>

The size of nanoparticles makes them optimal to be delivered with CED. Penetration of nanoparticles through the extracellular matrix in the brain is possible due to the larger effective pore size of the extracellular matrix (50 nm).<sup>126</sup> CED of dextran-coated maghemite MNPs have recently been depicted by MRI in a normal rat brain model,<sup>127</sup> showing that these particles could be directly imaged and tracked. They also showed that increased viscosity of the infusate increased efficacy of delivery and reduced leak back.

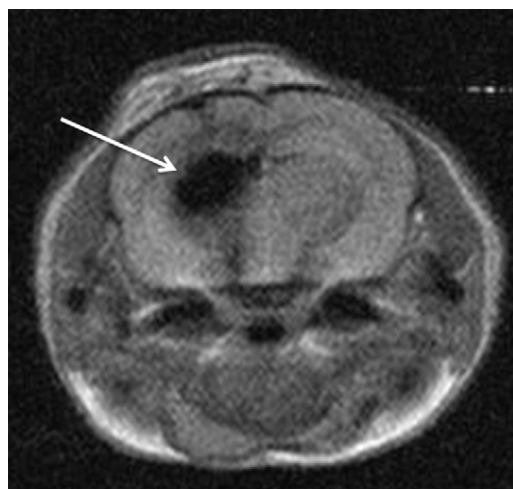
Imaging the infusate in CED is critical for ensuring adequate drug delivery to regions of interest. Valuable feedback can be gained from

tracking infusate delivered into the brain to enable clinicians to properly plan further treatments and avoid pitfalls, such as placement of catheters near sulci or ventricles.<sup>128,129</sup> Trials of conventional chemotherapeutics have failed to show significant benefit with CED, and lack of adequate drug delivery is often cited as the reason for this.<sup>130</sup> Although progress has been made using surrogate tracers, such as Gd-DTPA,<sup>131</sup> directly imaging the therapeutic particle would provide even more accurate information.

The authors have studied the CED of theranostic MNPs in mice (**Fig. 3**).<sup>35</sup> This particle consisted of an IONP core, coated by polymer and conjugated to an EGFR VIII antibody, specific for a subset of GBM tumors. The ability of the nanoparticles to localize to and image the lesion treated and its treatment effect were assessed. CED enabled a broad distribution of the nanoparticles in the region of the tumor and the surrounding brain, and repeat imaging showed that this effect remained for days after the nanoparticle delivery.

### **FUTURE STUDIES**

Although researchers have made great strides in developing nanoparticles that address the difficulties in treating GBM, many challenges remain. In the use of MNPs for thermotherapy and magnetic targeting, clinical equipment needs to be further developed and improved<sup>132</sup> to make these cost effective and freely available for further clinical trials. Phase III studies need to be undertaken to prove their effectiveness. In addition, drug delivery remains an issue with nanoparticles, and as further targeting motifs are studied, delivery of these



**Fig. 3.** CED of MNPs in the rodent brain. MRI of a rodent brain depicting the hypointense (white arrow) area in the brain that represents distribution of MNPs after CED with no leak back.

particles will be enhanced, further expanding their possible effectiveness.

## SUMMARY

Nanotechnology has quickly become a promising tool in the ongoing research to tackle the difficulties in treating GBM. The authors expect translational research to continue to elucidate further uses for this technology as these various particles come into widespread clinical use.

## REFERENCES

1. Brat DJ, Prayson RA, Ryken TC, et al. Diagnosis of malignant glioma: role of neuropathology. *J Neurooncol* 2008;89:287–311.
2. Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 2005;352:987–96.
3. Legler JM, Ries LA, Smith MA, et al. Cancer surveillance series [corrected]: brain and other central nervous system cancers: recent trends in incidence and mortality. *J Natl Cancer Inst* 1999;91:1382–90.
4. Brem H, Piantadosi S, Burger PC, et al. Placebo-controlled trial of safety and efficacy of intraoperative controlled delivery by biodegradable polymers of chemotherapy for recurrent gliomas. The Polymer-brain Tumor Treatment Group. *Lancet* 1995;345:1008–12.
5. Vredenburgh JJ, Desjardins A, Herndon JE 2nd, et al. Bevacizumab plus irinotecan in recurrent glioblastoma multiforme. *J Clin Oncol* 2007;25:4722–9.
6. Kelly PJ, Daumas-Duport C, Kispert DB, et al. Imaging-based stereotaxic serial biopsies in untreated intracranial glial neoplasms. *J Neurosurg* 1987;66:865–74.
7. Demuth T, Berens ME. Molecular mechanisms of glioma cell migration and invasion. *J Neurooncol* 2004;70:217–28.
8. Hayashi C, Ryozi U, Tasaki A. Ultra-fine particles: exploratory science and technology. Westwood (NJ): Noyes Publications; 1997. p. 2.
9. Maier-Hauff K, Ulrich F, Nestler D, et al. Efficacy and safety of intratumoral thermotherapy using magnetic iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent glioblastoma multiforme. *J Neurooncol* 2011;103(2):317–24.
10. Liu L, Venkatraman SS, Yang YY, et al. Polymeric micelles anchored with TAT for delivery of antibiotics across the blood-brain barrier. *Biopolymers* 2008;90:617–23.
11. Loo C, Lin A, Hirsch L, et al. Nanoshell-enabled photonics-based imaging and therapy of cancer. *Technol Cancer Res Treat* 2004;3:33–40.
12. Xing Y, Chaudry Q, Shen C, et al. Bioconjugated quantum dots for multiplexed and quantitative immunohistochemistry. *Nat Protoc* 2007;2:1152–65.
13. Provenzale JM, Silva GA. Uses of nanoparticles for central nervous system imaging and therapy. *AJNR Am J Neuroradiol* 2009;30:1293–301.
14. Jain TK, Richey J, Strand M, et al. Magnetic nanoparticles with dual functional properties: drug delivery and magnetic resonance imaging. *Biomaterials* 2008;29:4012–21.
15. Sun C, Lee JS, Zhang M. Magnetic nanoparticles in MR imaging and drug delivery. *Adv Drug Deliv Rev* 2008;60:1252–65.
16. Corot C, Robert P, Idee JM, et al. Recent advances in iron oxide nanocrystal technology for medical imaging. *Adv Drug Deliv Rev* 2006;58:1471–504.
17. Lodhia J, Mandarano G, Ferris N, et al. Development and use of iron oxide nanoparticles (Part 1): synthesis of iron oxide nanoparticles for MRI. *Biomed Imaging Interv J* 2010;6:e12.
18. Thorek DL, Chen AK, Czupryna J, et al. Superparamagnetic iron oxide nanoparticle probes for molecular imaging. *Ann Biomed Eng* 2006;34:23–38.
19. Weissleder R, Elizondo G, Wittenberg J, et al. Ultra-small superparamagnetic iron oxide: characterization of a new class of contrast agents for MR imaging. *Radiology* 1990;175:489–93.
20. Pan D, Caruthers SD, Hu G, et al. Ligand-directed nanobialys as theranostic agent for drug delivery and manganese-based magnetic resonance imaging of vascular targets. *J Am Chem Soc* 2008;130:9186–7.
21. Na HB, Lee JH, An K, et al. Development of a T1 contrast agent for magnetic resonance imaging using MnO nanoparticles. *Angew Chem Int Ed Engl* 2007;46:5397–401.
22. Bridot JL, Faure AC, Laurent S, et al. Hybrid gadolinium oxide nanoparticles: multimodal contrast agents for in vivo imaging. *J Am Chem Soc* 2007;129:5076–84.
23. Bourrinet P, Bengel HH, Bonnemain B, et al. Preclinical safety and pharmacokinetic profile of ferumoxtran-10, an ultrasmall superparamagnetic iron oxide magnetic resonance contrast agent. *Invest Radiol* 2006;41:313–24.
24. Aime S, Caravan P. Biodistribution of gadolinium-based contrast agents, including gadolinium deposition. *J Magn Reson Imaging* 2009;30:1259–67.
25. Abraham JL, Thakral C. Tissue distribution and kinetics of gadolinium and nephrogenic systemic fibrosis. *Eur J Radiol* 2008;66:200–7.
26. Varallyay P, Nesbit G, Muldoon LL, et al. Comparison of two superparamagnetic viral-sized iron oxide particles ferumoxides and ferumoxtran-10 with a gadolinium chelate in imaging intracranial tumors. *AJNR Am J Neuroradiol* 2002;23:510–9.

27. Neuwelt EA, Varallyay CG, Manninger S, et al. The potential of ferumoxytol nanoparticle magnetic resonance imaging, perfusion, and angiography in central nervous system malignancy: a pilot study. *Neurosurgery* 2007;60:601–11 [discussion: 611–2].
28. Neuwelt EA, Hamilton BE, Varallyay CG, et al. Ultra-small superparamagnetic iron oxides (USPIOs): a future alternative magnetic resonance (MR) contrast agent for patients at risk for nephrogenic systemic fibrosis (NSF)? *Kidney Int* 2009;75:465–74.
29. Moore A, Marecos E, Bogdanov A Jr, et al. Tumoral distribution of long-circulating dextran-coated iron oxide nanoparticles in a rodent model. *Radiology* 2000;214:568–74.
30. Zimmer C, Weissleder R, Poss K, et al. MR imaging of phagocytosis in experimental gliomas. *Radiology* 1995;197:533–8.
31. Villanueva A, Canete M, Roca AG, et al. The influence of surface functionalization on the enhanced internalization of magnetic nanoparticles in cancer cells. *Nanotechnology* 2009;20:115103.
32. Rhyner MN, Smith AM, Gao X, et al. Quantum dots and multifunctional nanoparticles: new contrast agents for tumor imaging. *Nanomedicine (Lond)* 2006;1:209–17.
33. Peng XH, Qian X, Mao H, et al. Targeted magnetic iron oxide nanoparticles for tumor imaging and therapy. *Int J Nanomedicine* 2008;3:311–21.
34. Remsen LG, McCormick CI, Roman-Goldstein S, et al. MR of carcinoma-specific monoclonal antibody conjugated to monocrySTALLINE iron oxide nanoparticles: the potential for noninvasive diagnosis. *AJNR Am J Neuroradiol* 1996;17:411–8.
35. Hadjipanayis CG, Machaidze R, Kaluzova M, et al. EGFRvIII antibody-conjugated iron oxide nanoparticles for magnetic resonance imaging-guided convection-enhanced delivery and targeted therapy of glioblastoma. *Cancer Res* 2010;70:6303–12.
36. Soroceanu L, Gillespie Y, Khazaeli MB, et al. Use of chlorotoxin for targeting of primary brain tumors. *Cancer Res* 1998;58:4871–9.
37. Lyons SA, O'Neal J, Sontheimer H. Chlorotoxin, a scorpion-derived peptide, specifically binds to gliomas and tumors of neuroectodermal origin. *Glia* 2002;39:162–73.
38. Deshane J, Garner CC, Sontheimer H. Chlorotoxin inhibits glioma cell invasion via matrix metalloproteinase-2. *J Biol Chem* 2003;278:4135–44.
39. Veisheh O, Gunn JW, Kievit FM, et al. Inhibition of tumor-cell invasion with chlorotoxin-bound superparamagnetic nanoparticles. *Small* 2009;5:256–64.
40. Veisheh O, Sun C, Fang C, et al. Specific targeting of brain tumors with an optical/magnetic resonance imaging nanoprobe across the blood-brain barrier. *Cancer Res* 2009;69:6200–7.
41. Veisheh O, Sun C, Gunn J, et al. Optical and MRI multifunctional nanoprobe for targeting gliomas. *Nano Lett* 2005;5:1003–8.
42. McFerrin MB, Sontheimer H. A role for ion channels in glioma cell invasion. *Neuron Glia Biol* 2006;2:39–49.
43. Christian S, Pilch J, Akerman ME, et al. Nucleolin expressed at the cell surface is a marker of endothelial cells in angiogenic blood vessels. *J Cell Biol* 2003;163:871–8.
44. Reddy GR, Bhojani MS, McConville P, et al. Vascular targeted nanoparticles for imaging and treatment of brain tumors. *Clin Cancer Res* 2006;12:6677–86.
45. Tomanek B, Iqbal U, Blasiak B, et al. Evaluation of brain tumor vessels specific contrast agents for glioblastoma imaging. *Neuro Oncol* 2012;14:53–63.
46. Faucher L, Guay-Begin AA, Lagueux J, et al. Ultra-small gadolinium oxide nanoparticles to image brain cancer cells in vivo with MRI. *Contrast Media Mol Imaging* 2011;6:209–18.
47. Mowat P, Mignot A, Rima W, et al. In vitro radiosensitizing effects of ultrasmall gadolinium based particles on tumour cells. *J Nanosci Nanotechnol* 2011;11:7833–9.
48. Fillmore HL, Shultz MD, Henderson SC, et al. Conjugation of functionalized gadolinium metallofullerenes with IL-13 peptides for targeting and imaging glial tumors. *Nanomedicine (Lond)* 2011;6:449–58.
49. Leung K. TAMRA-IL-13-Conjugated functionalized gadolinium metallofullerene (Gd<sub>3</sub>N@C<sub>80</sub>(OH)-26(CH<sub>2</sub>CH<sub>2</sub>COOH)-16), Molecular Imaging and Contrast Agent Database (MICAD). Bethesda (MD): National Library of Medicine (US), NCBI; 2004–2009. Available at: <http://micad.nih.gov>. Accessed April 27, 2012.
50. Lu YJ, Wei KC, Ma CC, et al. Dual targeted delivery of doxorubicin to cancer cells using folate-conjugated magnetic multi-walled carbon nanotubes. *Colloids Surf B Biointerfaces* 2012;89:1–9.
51. Sanai N, Polley MY, McDermott MW, et al. An extent of resection threshold for newly diagnosed glioblastomas. *J Neurosurg* 2011;115:3–8.
52. Senft C, Franz K, Blasel S, et al. Influence of iMRI-guidance on the extent of resection and survival of patients with glioblastoma multiforme. *Technol Cancer Res Treat* 2010;9:339–46.
53. Mehdorn HM, Schwartz F, Dawirs S, et al. High-field iMRI in glioblastoma surgery: improvement of resection radicality and survival for the patient? *Acta Neurochir Suppl* 2011;109:103–6.
54. Willems PW, Taphoorn MJ, Burger H, et al. Effectiveness of neuronavigation in resecting solitary intracerebral contrast-enhancing tumors: a randomized controlled trial. *J Neurosurg* 2006;104:360–8.
55. Hadjipanayis CG, Jiang H, Roberts DW, et al. Current and future clinical applications for optical imaging of cancer: from intraoperative surgical



- guidance to cancer screening. *Semin Oncol* 2011; 38:109–18.
56. Van Meir EG, Hadjipanayis CG, Norden AD, et al. Exciting new advances in neuro-oncology: the avenue to a cure for malignant glioma. *CA Cancer J Clin* 2010;60:166–93.
  57. Moore GE, Peyton WT, French LA, et al. The clinical use of fluorescein in neurosurgery; the localization of brain tumors. *J Neurosurg* 1948;5:392–8.
  58. Britz GW, Ghatan S, Spence AM, et al. Intracarotid RMP-7 enhanced indocyanine green staining of tumors in a rat glioma model. *J Neurooncol* 2002; 56:227–32.
  59. Ozawa T, Britz GW, Kinder DH, et al. Bromophenol blue staining of tumors in a rat glioma model. *Neurosurgery* 2005;57:1041–7 [discussion: 1041–7].
  60. Stummer W, Pichlmeier U, Meinel T, et al. Fluorescence-guided surgery with 5-aminolevulinic acid for resection of malignant glioma: a randomised controlled multicentre phase III trial. *Lancet Oncol* 2006;7:392–401.
  61. Eljamel MS, Goodman C, Moseley H. ALA and Photofrin fluorescence-guided resection and repetitive PDT in glioblastoma multiforme: a single centre Phase III randomised controlled trial. *Lasers Med Sci* 2008;23:361–7.
  62. Kircher MF, Mahmood U, King RS, et al. A multimodal nanoparticle for preoperative magnetic resonance imaging and intraoperative optical brain tumor delineation. *Cancer Res* 2003;63:8122–5.
  63. Aboody KS, Brown A, Rainov NG, et al. Neural stem cells display extensive tropism for pathology in adult brain: evidence from intracranial gliomas. *Proc Natl Acad Sci U S A* 2000;97:12846–51.
  64. Yang SY, Liu H, Zhang JN. Gene therapy of rat malignant gliomas using neural stem cells expressing IL-12. *DNA Cell Biol* 2004;23:381–9.
  65. Benedetti S, Pirola B, Pollo B, et al. Gene therapy of experimental brain tumors using neural progenitor cells. *Nat Med* 2000;6:447–50.
  66. Hamada H, Kobune M, Nakamura K, et al. Mesenchymal stem cells (MSC) as therapeutic cytoreagents for gene therapy. *Cancer Sci* 2005;96: 149–56.
  67. Wu X, Hu J, Zhou L, et al. In vivo tracking of superparamagnetic iron oxide nanoparticle-labeled mesenchymal stem cell tropism to malignant gliomas using magnetic resonance imaging. Laboratory investigation. *J Neurosurg* 2008;108:320–9.
  68. Tang C, Russell PJ, Martiniello-Wilks R, et al. Concise review: Nanoparticles and cellular carriers-allies in cancer imaging and cellular gene therapy? *Stem Cells* 2010;28:1686–702.
  69. Arbab AS, Janic B, Knight RA, et al. Detection of migration of locally implanted AC133+ stem cells by cellular magnetic resonance imaging with histological findings. *FASEB J* 2008;22:3234–46.
  70. Goldstein LS, Dewhirst MW, Repacholi M, et al. Summary, conclusions and recommendations: adverse temperature levels in the human body. *Int J Hyperthermia* 2003;19:373–84.
  71. Hildebrandt B, Wust P, Ahlers O, et al. The cellular and molecular basis of hyperthermia. *Crit Rev Oncol Hematol* 2002;43:33–56.
  72. Wust P, Hildebrandt B, Sreenivasa G, et al. Hyperthermia in combined treatment of cancer. *Lancet Oncol* 2002;3:487–97.
  73. Suto R, Srivastava PK. A mechanism for the specific immunogenicity of heat-shock protein-chaperoned peptides. *Science* 1995;269:1585–8.
  74. Santos-Marques MJ, Carvalho F, Sousa C, et al. Cytotoxicity and cell signalling induced by continuous mild hyperthermia in freshly isolated mouse hepatocytes. *Toxicology* 2006;224:210–8.
  75. Vanpouille-Box C, Lacoëuille F, Belloche C, et al. Tumor eradication in rat glioma and bypass of immunosuppressive barriers using internal radiation with (188)Re-lipid nanocapsules. *Biomaterials* 2011;32:6781–90.
  76. Hirsch LR, Gobin AM, Lowery AR, et al. Metal nanoshells. *Ann Biomed Eng* 2006;34:15–22.
  77. Bernardi RJ, Lowery AR, Thompson PA, et al. Immunonanoshells for targeted photothermal ablation in medulloblastoma and glioma: an in vitro evaluation using human cell lines. *J Neurooncol* 2008;86: 165–72.
  78. Baek SK, Makkouk AR, Krasieva T, et al. Photothermal treatment of glioma; an in vitro study of macrophage-mediated delivery of gold nanoshells. *J Neurooncol* 2011;104:439–48.
  79. Hadjipanayis CG, Bonder MJ, Balakrishnan S, et al. Metallic iron nanoparticles for MRI contrast enhancement and local hyperthermia. *Small* 2008;4:1925–9.
  80. Mehdaoui B, Meffre A, Carrey J, et al. Optimal size of nanoparticles for magnetic hyperthermia: a combined theoretical and experimental study. *Adv Funct Mater* 2011;21:4573–81.
  81. Dennis CL, Jackson AJ, Borchers JA, et al. Nearly complete regression of tumors via collective behavior of magnetic nanoparticles in hyperthermia. *Nanotechnology* 2009;20:395103.
  82. Lee JH, Jang JT, Choi JS, et al. Exchange-coupled magnetic nanoparticles for efficient heat induction. *Nat Nanotechnol* 2011;6:418–22.
  83. Wijaya A, Brown KA, Alper JD, et al. Magnetic field heating study of Fe-doped Au nanoparticles. *J Magn Magn Mater* 2007;309:15–9.
  84. Sharma R, Chen CJ. Newer nanoparticles in hyperthermia treatment and thermometry. *J Nanopart Res* 2009;11:671–89.
  85. Pradhan P, Giri J, Samanta G, et al. Comparative evaluation of heating ability and biocompatibility of different ferrite-based magnetic fluids for

- hyperthermia application. *J Biomed Mater Res B Appl Biomater* 2007;81:12–22.
86. Kim DH, Thai YT, Nikles DE, et al. Heating of aqueous dispersions containing MnFe(2)O(4) nanoparticles by radio-frequency magnetic field induction. *IEEE Trans Magn* 2009;45:64–70.
  87. Kaman O, Pollert E, Veverka P, et al. Silica encapsulated manganese perovskite nanoparticles for magnetically induced hyperthermia without the risk of overheating. *Nanotechnology* 2009;20:275610.
  88. Atsarkin VA, Levkin LV, Posvyanskiy VS, et al. Solution to the bioheat equation for hyperthermia with La1-xAgyMnO3-nanoparticles: the effect of temperature autostabilization. *Int J Hyperthermia* 2009;25:240–7.
  89. Bae S, Lee SW, Takemura Y, et al. Dependence of frequency and magnetic field on self-heating characteristics of NiFe2O4 nanoparticles for hyperthermia. *IEEE Trans Magn* 2006;42:3566–8.
  90. Kim DH, Lee SH, Kim KN, et al. Temperature change of various ferrite particles with alternating magnetic field for hyperthermic application. *J Magn Magn Mater* 2005;293:320–7.
  91. Kim DH, Lee SH, Kim KN, et al. In vitro and in vivo characterization of various ferrites for hyperthermia in cancer-treatment. In: Li P, Zhang K, Colwell CW Jr, editors. *Bioceramics*, vol. 17. (Key Engineering Materials) Switzerland: Trans Tech Publications; 2005. p. 827–30.
  92. Huber DL. Synthesis, properties, and applications of iron nanoparticles. *Small* 2005;1:482–501.
  93. Pradhan P, Giri J, Banerjee R, et al. Cellular interactions of lauric acid and dextran-coated magnetite nanoparticles. *J Magn Magn Mater* 2007;311:282–7.
  94. Luis Corchero J, Villaverde A. Biomedical applications of distally controlled magnetic nanoparticles. *Trends Biotechnol* 2009;27:468–76.
  95. Jordan A, Scholz R, Maier-Hauff K, et al. The effect of thermotherapy using magnetic nanoparticles on rat malignant glioma. *J Neurooncol* 2006;78:7–14.
  96. Maier-Hauff K, Rothe R, Scholz R, et al. Intracranial thermotherapy using magnetic nanoparticles combined with external beam radiotherapy: results of a feasibility study on patients with glioblastoma multiforme. *J Neurooncol* 2007;81:53–60.
  97. Jiang X, Xin H, Sha X, et al. PEGylated poly(trimethylene carbonate) nanoparticles loaded with paclitaxel for the treatment of advanced glioma: in vitro and in vivo evaluation. *Int J Pharm* 2011;420:385–94.
  98. Sawyer AJ, Saucier-Sawyer JK, Booth CJ, et al. Convection-enhanced delivery of camptothecin-loaded polymer nanoparticles for treatment of intracranial tumors. *Drug Deliv Transl Res* 2011;1:34–42.
  99. Agarwal A, Mackey MA, El-Sayed MA, et al. Remote triggered release of doxorubicin in tumors by synergistic application of thermosensitive liposomes and gold nanorods. *ACS Nano* 2011;5:4919–26.
  100. Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* 2008;455:1061–8.
  101. Jin J, Bae KH, Yang H, et al. In vivo specific delivery of c-Met siRNA to glioblastoma using cationic solid lipid nanoparticles. *Bioconjug Chem* 2011;22:2568–72.
  102. Zhan C, Meng Q, Li Q, et al. Cyclic RGD-polyethylene glycol-polyethylenimine for intracranial glioblastoma-targeted gene delivery. *Chem Asian J* 2012;7:91–6.
  103. Shultz MD, Wilson JD, Fuller CE, et al. Metallofullerene-based nanopatform for brain tumor brachytherapy and longitudinal imaging in a murine orthotopic xenograft model. *Radiology* 2011;261:136–43.
  104. Nie S, Xing Y, Kim GJ, et al. Nanotechnology applications in cancer. *Annu Rev Biomed Eng* 2007;9:257–88.
  105. Peer D, Karp JM, Hong S, et al. Nanocarriers as an emerging platform for cancer therapy. *Nat Nanotechnol* 2007;2:751–60.
  106. Gref R, Minamitake Y, Peracchia MT, et al. Biodegradable long-circulating polymeric nanospheres. *Science* 1994;263:1600–3.
  107. van der Sanden BP, Rozijn TH, Rijken PF, et al. Noninvasive assessment of the functional neovasculature in 9L-glioma growing in rat brain by dynamic 1H magnetic resonance imaging of gadolinium uptake. *J Cereb Blood Flow Metab* 2000;20:861–70.
  108. Vajkoczy P, Menger MD. Vascular microenvironment in gliomas. *J Neurooncol* 2000;50:99–108.
  109. Batchelor TT, Sorensen AG, di Tomaso E, et al. AZD2171, a pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients. *Cancer Cell* 2007;11:83–95.
  110. Son YJ, Jang JS, Cho YW, et al. Biodistribution and anti-tumor efficacy of doxorubicin loaded glycol-chitosan nanoaggregates by EPR effect. *J Control Release* 2003;91:135–45.
  111. Agemy L, Friedmann-Morvinski D, Kotamraju VR, et al. Targeted nanoparticle enhanced proapoptotic peptide as potential therapy for glioblastoma. *Proc Natl Acad Sci U S A* 2011;108:17450–5.
  112. Li J, Gu B, Meng Q, et al. The use of myristic acid as a ligand of polyethylenimine/DNA nanoparticles for targeted gene therapy of glioblastoma. *Nanotechnology* 2011;22:435101.
  113. Tahara K, Kato Y, Yamamoto H, et al. Intracellular drug delivery using polysorbate 80-modified poly(D, L-lactide-co-glycolide) nanospheres to glioblastoma cells. *J Microencapsul* 2011;28:29–36.

114. Gelperina S, Maksimenko O, Khalansky A, et al. Drug delivery to the brain using surfactant-coated poly(lactide-co-glycolide) nanoparticles: influence of the formulation parameters. *Eur J Pharm Biopharm* 2010;74:157–63.
115. Wohlfart S, Khalansky AS, Gelperina S, et al. Efficient chemotherapy of rat glioblastoma using doxorubicin-loaded PLGA nanoparticles with different stabilizers. *PLoS One* 2011;6:e19121.
116. Chang J, Paillard A, Passirani C, et al. Transferrin adsorption onto PLGA nanoparticles governs their interaction with biological systems from blood circulation to brain cancer cells. *Pharm Res* 2011;29:1495–505.
117. Chertok B, Moffat BA, David AE, et al. Iron oxide nanoparticles as a drug delivery vehicle for MRI monitored magnetic targeting of brain tumors. *Biomaterials* 2008;29:487–96.
118. Chertok B, David AE, Huang Y, et al. Glioma selectivity of magnetically targeted nanoparticles: a role of abnormal tumor hydrodynamics. *J Control Release* 2007;122:315–23.
119. Pulfer SK, Ciccotto SL, Gallo JM. Distribution of small magnetic particles in brain tumor-bearing rats. *J Neurooncol* 1999;41:99–105.
120. Hynynen K, McDannold N, Vykhodtseva N, et al. Focal disruption of the blood-brain barrier due to 260-kHz ultrasound bursts: a method for molecular imaging and targeted drug delivery. *J Neurosurg* 2006;105:445–54.
121. Pardridge WM. Drug and gene delivery to the brain: the vascular route. *Neuron* 2002;36:555–8.
122. Muldoon LL, Soussain C, Jahnke K, et al. Chemotherapy delivery issues in central nervous system malignancy: a reality check. *J Clin Oncol* 2007;25:2295–305.
123. Liu HL, Hua MY, Yang HW, et al. Magnetic resonance monitoring of focused ultrasound/magnetic nanoparticle targeting delivery of therapeutic agents to the brain. *Proc Natl Acad Sci U S A* 2010;107:15205–10.
124. Bobo RH, Laske DW, Akbasak A, et al. Convection-enhanced delivery of macromolecules in the brain. *Proc Natl Acad Sci U S A* 1994;91:2076–80.
125. Allard E, Passirani C, Benoit JP. Convection-enhanced delivery of nanocarriers for the treatment of brain tumors. *Biomaterials* 2009;30:2302–18.
126. Thorne RG, Nicholson C. In vivo diffusion analysis with quantum dots and dextrans predicts the width of brain extracellular space. *Proc Natl Acad Sci U S A* 2006;103:5567–72.
127. Perlstein B, Ram Z, Daniels D, et al. Convection-enhanced delivery of maghemite nanoparticles: increased efficacy and MRI monitoring. *Neuro Oncol* 2008;10:153–61.
128. Sampson JH, Brady ML, Petry NA, et al. Intracerebral infusate distribution by convection-enhanced delivery in humans with malignant gliomas: descriptive effects of target anatomy and catheter positioning. *Neurosurgery* 2007;60:ONS89–98 [discussion: ONS98–9].
129. Varenika V, Dickinson P, Bringas J, et al. Detection of infusate leakage in the brain using real-time imaging of convection-enhanced delivery. *J Neurosurg* 2008;109:874–80.
130. Sampson JH, Archer G, Pedain C, et al. Poor drug distribution as a possible explanation for the results of the PRECISE trial. *J Neurosurg* 2010;113:301–9.
131. Asthagiri AR, Walbridge S, Heiss JD, et al. Effect of concentration on the accuracy of convective imaging distribution of a gadolinium-based surrogate tracer. *J Neurosurg* 2011;115:467–73.
132. Silva AC, Oliveira TR, Mamani JB, et al. Application of hyperthermia induced by superparamagnetic iron oxide nanoparticles in glioma treatment. *Int J Nanomedicine* 2011;6:591–603.